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# The Potential Effectiveness of Probiotic-Based Sanitation Procedures in Nosocomial Infection Control: A Review Article



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## Abstract

The sanitation of the hospital environment for the purpose of preventing the transmission of nosocomial infections has a major role in reducing the infection of hospitalized patients with the bacteria living on hospital surfaces. The excessive use of chemical detergents in recent decades has led to microbial resistance in nosocomial infectious bacteria. Researchers' attention has therefore been drawn to the use of probiotics for disinfecting hospital surfaces. The present study was conducted to assess the potential effectiveness of probiotic products in controlling the contamination of inert surfaces in the environment and medical instruments in health centers and investigate the claim that the antagonistic property of probiotic microorganisms offers an effective method for controlling nosocomial infections and a suitable alternative to conventional disinfection methods. A search was carried out for relevant articles published from 2000 to 2018 in databases including ISI, PubMed, Scopus, EMBASE, and Google Scholar, using the keywords "nosocomial infections", "disinfection", "sanitation", "probiotics" and "infected surfaces". The articles published from 2000 to 2018 confirmed the greater effectiveness of probiotic disinfection (by up to 90%) compared to conventional chemical disinfection in controlling nosocomial infections. Nevertheless, more extensive studies are needed on probiotics to determine the possibility of replacing good bacteria with bad bacteria in future decades.

Keywords: Nosocomial infection, Disinfection, Probiotic, Infected surfaces, Pathogens

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#### 1. Introduction

Contaminated surfaces in hospitals provide an important source for transmission of health careassociated pathogens (1). Therefore, the use of disinfectants has a major role in the management of nosocomial infections. As a huge challenge throughout the world, nosocomial infections threaten the health of 15% of hospitalized patients (2,3). Pathogens including vancomycin-resistant Enterococcus, methicillin-resistant Staphylococcus aureus, Norovirus, Clostridium difficile, and multidrug-resistant gram-negative bacteria (MDR) can survive in the hospital environment for days and months and act as a source of spreading infection (4-6). Conventional sanitation based on the use of chemicals not only is ineffective in preventing the re-infection of surfaces but also produces multidrug-resistant microbial strains. The theory of probiotic-based improvement was formed and probiotic microorganisms were first introduced by Kramer et al, who pointed out their antagonistic effect

on the proliferation of pathogens on hospital surfaces (3). By producing the extracellular polysaccharide matrix, Staphylococcus aureus and Escherichia coli adhere to inert surfaces and produce biofilm (7). This biofilm formation enables microorganisms to benefit from survival against their planktonic (free-floating) rivals and provides an optimal environment for the growth, proliferation, gene transfer, and the microbial population development. The proximity of rival species can have an antagonistic effect on the formation of biofilm (8). Lactobacillus species can produce amphipathic molecules called bio-surfactant, which have anti-adhesion properties against pathogenic microbes (9,10). The extensive use of chemical disinfectants not only produces disinfectantresistant microbial strains but also makes them resistant against antibiotics. According to a recent report, using chlorhexidine leads to the emergence of multidrugresistant gram-negative bacteria (MDR-GNB) against colistin. This antibiotic was used until 2016 as the drug of

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last resort for the treatment of infections caused by MDR-GNB (11). The potential to cause microbial resistance is a highly undesirable side-effect of chemical detergents, and the spread of pathogens is responsible for a large part of nosocomial infections (12,13). Self-disinfecting method can be proposed as an alternative method in which hospital surfaces are smeared with a bactericidal agent using silver and copper. This method is expensive and not suitable for all surfaces (14). The issue of minimizing the risk of nosocomial infection and preventing the increase in drug resistance has turned the attention of researchers to the management of "hospital environment sanitation" instead of "patients' physical health" (15,16). The present article aims to review the potential effectiveness of biosurfactants in controlling nosocomial infections in studies conducted from 2000 to 2018.

# 2. Materials and Methods

# 2.1. Study Objective

The present study aimed to review articles published from 2000 to 2018 to find evidence in favor of the claim that probiotics have the required effectiveness for controlling nosocomial infections.

## 2.2. Study Strategy

Published literature was retrieved through a search using the words (MeSH was applied to select appropriate keywords) "microbial infection", "infection control", "disinfectant", "probiotic organisms", "bio-surfactant", "nosocomial infections" and "sanitation" in databases including PubMed, EMBASE, ISI, and Google Scholar (2000-2018) (Fig. 1).

## 2.3. Article Inclusion Criteria and Data Extraction

First, the abstracts of all the retrieved papers were reviewed to select the relevant articles. Then, data were extracted by reviewing the full text of the selected articles. The data extraction criteria consisted of the type of experiment, results, type of probiotic and type of hospital organism.

#### 3. Results and Discussion

Of the 65 articles retrieved, 15 were identified as relevant (Fig. 1). After printing out the full text of all these articles, the careful assessment of the data and results was carried out personally. Half of the experiments were in vitro experiments and the other half were field evidence. The *Bacillus* species made up a greater proportion of the probiotic bacteria. According to Table 1, the probiotic-based sanitation methods used in the experiments had been able to reduce pathogens on hospital surfaces about 50% to 100% more than conventional chemical methods. It is a generally-accepted fact that some good bacteria are crucial and valuable for human health and can be effectively used in the prevention and treatment of infectious diseases (32). Fig. 2 shows the schematic plan of replacing pathogens with probiotic bacteria.

Health care-associated infections are one of the serious problems concerning the safety of public health worldwide. As a challenge throughout the world, nosocomial infections threaten the health of 15% of hospitalized patients (33). It is well known that contaminated surfaces in hospitals act as reservoirs for the pathogenic bacteria, increasing the risk of infection (34). The extensive use of the traditional chemical disinfectants



Fig.1. Flow Chart of Literature Review (2000-2018) of Included and Excluded Studies for Nosocomial Disinfection by Probiotics.



**Fig. 2.** The Schematic Plan of the Antagonistic Effect of Probiotic Bacteria on Pathogens Living on Hospital Surfaces and Medical Instruments.

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	Method	Reference	Probiotics	Surfaces	Hospital Bacteria	Outcome
•	In situ	Caselli et al (2018) (17)	PCHS (probiotic cleaning hygiene system)	Floor, Sink, Footboard	Staphylococcus spp., Enterobacteriaceae spp., Acinetobacter spp., Mycetes, Pseudomonas spp., Clostridium difficile	Over 83% reduction of surfaces pathogens
	Commercial hatchery	Graham et al (2018) (18)	Bacillus subtilis		Staphylococcus, Pseudomonas, Escherichia coli	
	In situ	Caselli (2017) (19)	Bacillus subtilis, Bacillus pumilus, Bacillus megaterium	Floor, Sink	Common bacteria	Over 90% reduction compared to the routine methods
	In situ	Caselli et al (2016) (20)	Controlled pre-post interventional study		Hospital bacteria	Over 90% reduction compared to the routine methods
	In vitro, in situ (opinion)	Caselli et al (2016) (21)	Bacillus subtilis, Bacillus pumilus, Bacillus megaterium			Over 90% reduction compared to the routine methods
	In vitro	Fanci et al (2015) (22)		Floor, Washbasin, Desk	Enterococcus faecalis, Candida albicans, Pseudomonas, Acinetobacter, Klebsiella pneumoniae	92.2%-99.9% bacterial reduction, 100% reduction of <i>Pseudomonas, Acinetobacter</i>
	In vitro	Vandini et al (2014) (23)	Bacillus subtilis spp. spores	Floor, Sink, Toilet	Staphylococcus aureus Coliforms Pseudomonas spp. Candida spp.	Over 80% reduction of microbial load
	In situ	Vandini et al (2014) (24)	Genus of bacillus	Floor, Doors,Showers toilet, Windows sill, Sink (all made of stone, plastic, wood, glass, and metal)	Coliforms, Staphylococcus aureus, Clostridium difficile, Candida albicans	50%-89% reduction in microbial load on the surfaces
	In situ and in vitro	Mazzacane et al (2014) (25)	Genus of <i>Bacillus</i> , vegetative form and spore	Toilet, Floor, Corridor floor	Coliforms, Staphylococcus aureus, Candida albicans, Acinetobacter spp., Pseudomonas spp.	About 90% reduction in microbial cell
	In vitro	Walencke et al (2008) (26)	Lactobacillus acidophilus	Floor	Staphylococcus aureus, Staphylococcus epidermidis biofilm	About 5%-56% reduction
	In vitro	Rodrigues et al (2006) (27)	Streptococcus thermophilus	Silicon, Rubber	Coliforms, Staphylococcus aureus, Candida albicans, Acinetobacter spp., Pseudomonas spp.	97% reduction in adhesion of bacterial cell
	In vitro	Rodrigues et al (2004) (28)	Lactococcus lactis	Silicon, Rubber	Staphylococcus aureus, Candida albicans, Staphylococcus epidermidis, Streptococcus salivarius	Adhesion of pathogenic bacteria
	In vitro	Rodrigues et al (2004) (29)	Streptococcus thermophilus, Lactococcus lactis	Silicon, Rubber	Staphylococcus aureus, Candida albicans, Staphylococcus epidermidis, Streptococcus salivarius	Significant decrease in the number of bacteria on the surface
	In vitro	Hoogmoed et al (30)	Streptococcus mitis	Glass	Streptococcus mutans	Release of biosurfactant by Streptococcus mitis and decrease in the number of s. mutans cells
	In vitro	Vander Mei et al (2000) (31)	Lactobacillus spp., Streptococcus thermophilus	Silicon, Rubber	Candida spp., Streptococcus spp., Staphylococcus spp.	Decrease in the number of pathogens on the surface

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not only produces disinfectant-resistant microbial strains but also makes them resistant against antibiotics (11). The issue of minimizing the risk of nosocomial infection and the safety of public health has turned the attention of the researchers to the use of probiotic bacteria (17-31).

The results of the present review study show that despite the significant results reported, which generally confirm the effect of the probiotic-based sanitation method, a definitive answer cannot yet be given to the following questions: 1) How do probiotic microorganisms survive on inert surfaces and how long can they keep up and continue their dominance in competition with hospital pathogens? 2) To what extent are probiotics able to fight and act as antagonists against pathogenic microorganism growth? 3) How can we ensure the safety of probiotics and probiotic-containing detergents for consistent use in hospitals? 4) How can hospitalized patients' health be promoted as a result of shifting from pathogens to probiotics? In other words, to what extent does replacing nosocomial pathogens with probiotics have a share in preventing dangerous pulmonary, gastric and urinary infections in patients? 5) Are probiotics and their products enough for inhibiting all microbial species? and 6) Is it possible for the epidemiology of nosocomial infections to be shifted toward microorganisms on which probiotics have no effect?

The in vitro experiments conducted on hydrophobic materials (such as glass) and hydrophilic materials (such as silicon) somewhat support the first question (35). The safety of using the probiotic-based sanitation method has been confirmed by the European Union Workshop (36), although several species of lactobacillus used in humans as probiotics have reportedly caused lactobacillus bacteremia (a very rare disease) in patients with serious underlying diseases (37). There has been a reduction in the frequency of nosocomial infections, diarrhea, colon infections (38), upper respiratory tract infections (39), and surgical site infections (40). The environmental parameters, such as humidity, temperature, and microbial flora, of different hospital units are regarded as factors affecting the disinfection of hospital surfaces and should therefore be considered in all antimicrobial processes.

This review study shows that a probiotic-based cleaning strategy is more effective (up to 90%) in reducing microbe, compared to a traditional chemical disinfectant. However, to develop a better understanding of the mechanism of the effectiveness of probiotics, further studies are needed.

#### 4. Conclusion

The literature review (2000-2018) confirmed the greater effectiveness of probiotic disinfection (by up to 90%) compared to traditional chemical disinfection in controlling nosocomial infections. Since it is not possible to fully disinfect pathogens from all surfaces, nosocomial infections and drug-resistant pathogens can perhaps be controlled by relying on greater knowledge about the

defects and shortcomings of the idea of using probiotics and by replacing "bad bacteria" with "good bacteria". There is no easy way to remove microbes from surfaces. As a "green" alternative to chemical disinfectants, probiotics are biodegradable and environment-friendly. Using them is an innovative way to disinfect hospital environments effectively.

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#### References

- Dancer SJ. Importance of the environment in meticillinresistant Staphylococcus aureus acquisition: the case for hospital cleaning. Lancet Infect Dis. 2008;8(2):101-13. doi: 10.1016/s1473-3099(07)70241-4.
- Dancer SJ. The role of environmental cleaning in the control of hospital-acquired infection. J Hosp Infect. 2009;73(4):378-85. doi: 10.1016/j.jhin.2009.03.030.
- Kramer A, Schwebke I, Kampf G. How long do nosocomial pathogens persist on inanimate surfaces? a systematic review. BMC Infect Dis. 2006;6:130. doi: 10.1186/1471-2334-6-130.
- Wagenvoort JH, Sluijsmans W, Penders RJ. Better environmental survival of outbreak vs. sporadic MRSA isolates. J Hosp Infect. 2000;45(3):231-4. doi: 10.1053/ jhin.2000.0757.
- Chiang SR, Chuang YC, Tang HJ, Chen CC, Chen CH, Lee NY, et al. Intratracheal colistin sulfate for BALB/c mice with early pneumonia caused by carbapenem-resistant Acinetobacter baumannii. Crit Care Med. 2009;37(9):2590-5. doi: 10.1097/ CCM.0b013e3181a0f8e1.
- Lawley TD, Clare S, Deakin LJ, Goulding D, Yen JL, Raisen C, et al. Use of purified Clostridium difficile spores to facilitate evaluation of health care disinfection regimens. Appl Environ Microbiol. 2010;76(20):6895-900. doi: 10.1128/aem.00718-10.
- Weber DJ, Rutala WA, Miller MB, Huslage K, Sickbert-Bennett E. Role of hospital surfaces in the transmission of emerging health care-associated pathogens: norovirus, Clostridium difficile, and Acinetobacter species. Am J Infect Control. 2010;38(5 Suppl 1):S25-33. doi: 10.1016/j.ajic.2010.04.196.
- Schrezenmeir J, de Vrese M. Probiotics, prebiotics, and synbiotics--approaching a definition. Am J Clin Nutr. 2001;73(2 Suppl):361S-4S. doi: 10.1093/ajcn/73.2.361s.
- Santos VL, Nardi RM, Dias-Souza MV. Biosurfactants as Antimicrobial and Antibiofilm Agents. In: Current Developments in Biotechnology and Bioengineering. Amsterdam: Elsevier; 2017. p 371.
- Falagas ME, Rafailidis PI, Makris GC. Bacterial interference for the prevention and treatment of infections. Int J Antimicrob Agents. 2008;31(6):518-22. doi: 10.1016/j. ijantimicag.2008.01.024.
- Servin AL. Antagonistic activities of lactobacilli and bifidobacteria against microbial pathogens. FEMS Microbiol Rev. 2004;28(4):405-40. doi: 10.1016/j.femsre.2004.01.003.
- Singh P, Cameotra SS. Potential applications of microbial surfactants in biomedical sciences. Trends Biotechnol. 2004;22(3):142-6. doi: 10.1016/j.tibtech.2004.01.010.
- 13. Wand ME, Bock LJ, Bonney LC, Sutton JM. Mechanisms of

increased resistance to chlorhexidine and cross-resistance to colistin following exposure of Klebsiella pneumoniae clinical isolates to chlorhexidine. Antimicrob Agents Chemother. 2017;61(1). doi: 10.1128/aac.01162-16.

- Caini S, Hajdu A, Kurcz A, Böröcz K. Hospital-acquired infections due to multidrug-resistant organisms in Hungary, 2005-2010. Euro Surveill. 2013;18(2):20352.
- Cornejo-Juárez P, Vilar-Compte D, Pérez-Jiménez C, Ñamendys-Silva SA, Sandoval-Hernández S, Volkow-Fernández P. The impact of hospital-acquired infections with multidrug-resistant bacteria in an oncology intensive care unit. Int J Infect Dis. 2015;31:31-4. doi: 10.1016/j. ijid.2014.12.022.
- Otter JA, Yezli S, Perl TM, Barbut F, French GL. The role of 'no-touch' automated room disinfection systems in infection prevention and control. J Hosp Infect. 2013;83(1):1-13. doi: 10.1016/j.jhin.2012.10.002.
- Pettigrew MM, Johnson JK, Harris AD. The human microbiota: novel targets for hospital-acquired infections and antibiotic resistance. Ann Epidemiol. 2016;26(5):342-7. doi: 10.1016/j. annepidem.2016.02.007.
- Koenigsknecht MJ, Young VB. Faecal microbiota transplantation for the treatment of recurrent Clostridium difficile infection: current promise and future needs. Curr Opin Gastroenterol. 2013;29(6):628-32. doi: 10.1097/MOG.0b013e328365d326.
- Caselli E, Brusaferro S, Coccagna M, Arnoldo L, Berloco F, Antonioli P, et al. Reducing healthcare-associated infections incidence by a probiotic-based sanitation system: a multicentre, prospective, intervention study. PLoS One. 2018;13(7):e0199616. doi: 10.1371/journal.pone.0199616.
- Vandini A, Frabetti A, Antonioli P, Platano D, Branchini A, Camerada MT, et al. Reduction of the microbiological load on hospital surfaces through probiotic-based cleaning procedures: a new strategy to control nosocomial infections. J Microbiol Exp. 2014;1(5):153-61. doi: 10.15406/ jmen.2014.01.00027.
- Caselli E. Hygiene: microbial strategies to reduce pathogens and drug resistance in clinical settings. Microb Biotechnol. 2017;10(5):1079-83. doi: 10.1111/1751-7915.12755.
- La Fauci V, Costa GB, Anastasi F, Facciolà A, Grillo OC, Squeri R. An Innovative Approach to Hospital Sanitization Using Probiotics: In Vitro and Field Trials. J Microb Biochem Technol. 2015;7(3):160-4. doi: 10.4172/1948-5948.1000198.
- Vandini A, Temmerman R, Frabetti A, Caselli E, Antonioli P, Balboni PG, et al. Hard surface biocontrol in hospitals using microbial-based cleaning products. PLoS One. 2014;9(9):e108598. doi: 10.1371/journal.pone.0108598.
- Vandini A, Frabetti A, Antonioli P, Platano D, Branchini A, Camerada MT, et al. Reduction of the microbiological load on hospital surfaces through probiotic-based cleaning procedures: a new strategy to control nosocomial infections. J Microbiol Exp. 2014;1(5):153-61. doi: 10.15406/ jmen.2014.01.00027.
- 25. Mazzacane S, Finzi G, Aparo L, Balboni PG, Vandini A, Lanzoni L, et al. The sanitation of hospital stays: new strategies for the reduction of HAIs. Health Manage. 2014;14(3):1-12.
- Walencka E, Rózalska S, Sadowska B, Rózalska B. The influence of Lactobacillus acidophilus-derived surfactants on staphylococcal adhesion and biofilm formation. Folia Microbiol (Praha). 2008;53(1):61-6. doi: 10.1007/s12223-008-0009-y.

- 27. Rodrigues L, van der Mei H, Banat IM, Teixeira J, Oliveira R. Inhibition of microbial adhesion to silicone rubber treated with biosurfactant from Streptococcus thermophilus A. FEMS Immunol Med Microbiol. 2006;46(1):107-12. doi: 10.1111/j.1574-695X.2005.00006.x.
- 28. Rodrigues L, van der Mei H, Teixeira JA, Oliveira R. Biosurfactant from Lactococcus lactis 53 inhibits microbial adhesion on silicone rubber. Appl Microbiol Biotechnol. 2004;66(3):306-11. doi: 10.1007/s00253-004-1674-7.
- Rodrigues L, van der Mei HC, Teixeira J, Oliveira R. Influence of biosurfactants from probiotic bacteria on formation of biofilms on voice prostheses. Appl Environ Microbiol. 2004;70(7):4408-10. doi: 10.1128/aem.70.7.4408-4410.2004.
- 30. van Hoogmoed CG, van der Kuijl-Booij M, van der Mei HC, Busscher HJ. Inhibition of Streptococcus mutans NS adhesion to glass with and without a salivary conditioning film by biosurfactant- releasing Streptococcus mitis strains. Appl Environ Microbiol. 2000;66(2):659-63. doi: 10.1128/ aem.66.2.659-663.2000.
- 31. van der Mei HC, Free RH, Elving GJ, Van Weissenbruch R, Albers FW, Busscher HJ. Effect of probiotic bacteria on prevalence of yeasts in oropharyngeal biofilms on silicone rubber voice prostheses in vitro. J Med Microbiol. 2000;49(8):713-8. doi: 10.1099/0022-1317-49-8-713.
- Al-Ghalith GA, Knights D. Bygiene: the new paradigm of bidirectional hygiene. Yale J Biol Med. 2015;88(4):359-65.
- Burke JP. Infection control a problem for patient safety. N Engl J Med. 2003;348(7):651-6. doi: 10.1056/NEJMhpr020557.
- 34. European Food Safety Authority (EFSA). Opinion of the Scientific Committee on a request from EFSA related to a generic approach to the safety assessment by EFSA of microorganisms used in food/feed and the production of food/feed additives. EFSA J. 2005;3(6):226. doi: 10.2903/j. efsa.2005.226.
- 35. Lebeer S, Verhoeven TL, Perea Velez M, Vanderleyden J, De Keersmaecker SC. Impact of environmental and genetic factors on biofilm formation by the probiotic strain Lactobacillus rhamnosus GG. Appl Environ Microbiol. 2007;73(21):6768-75. doi: 10.1128/aem.01393-07.
- Adams MR, Marteau P. On the safety of lactic acid bacteria from food. Int J Food Microbiol. 1995;27(2-3):263-4. doi: 10.1016/0168-1605(95)00067-t.
- Husni RN, Gordon SM, Washington JA, Longworth DL. Lactobacillus bacteremia and endocarditis: review of 45 cases. Clin Infect Dis. 1997;25(5):1048-55. doi: 10.1086/516109.
- Giamarellos-Bourboulis EJ, Bengmark S, Kanellakopoulou K, Kotzampassi K. Pro- and synbiotics to control inflammation and infection in patients with multiple injuries. J Trauma. 2009;67(4):815-21. doi: 10.1097/TA.0b013e31819d979e.
- 39. Banupriya B, Biswal N, Srinivasaraghavan R, Narayanan P, Mandal J. Probiotic prophylaxis to prevent ventilator associated pneumonia (VAP) in children on mechanical ventilation: an open-label randomized controlled trial. Intensive Care Med. 2015;41(4):677-85. doi: 10.1007/s00134-015-3694-4.
- Sommacal HM, Bersch VP, Vitola SP, Osvaldt AB. Perioperative synbiotics decrease postoperative complications in periampullary neoplasms: a randomized, doubleblind clinical trial. Nutr Cancer. 2015;67(3):457-62. doi: 10.1080/01635581.2015.1004734.