

# Air Pollution, Ozone, and Sulfur Dioxide Can Affect the Blood Serum Lipid Profile and Oxidative Stress of Male Wistar Rats

Hossein Mashhadi-Abdolahi<sup>1</sup>, Roya Darbani<sup>2</sup>, Oldouz Rabet<sup>2</sup>, Asal Golchin<sup>3</sup>, Sorayya Kheirouri<sup>4</sup>, Mohammad Alizadeh<sup>5</sup>, Mehran Mesgari-Abbasi<sup>2\*</sup>

<sup>1</sup>Tabriz Health Services Management Research Center, Tabriz, Iran

<sup>2</sup>Drug Applied Research Center, Tabriz University of Medical Sciences, Tabriz, Iran

<sup>3</sup>Department of Clinical Biochemistry, Faculty of Medicine, Urmia University of Medical Sciences, Urmia, Iran

<sup>4</sup>Department of Nutrition, Tabriz University of Medical Sciences, Tabriz, Iran

<sup>5</sup>Nutrition Research Center, Tabriz University of Medical Sciences, Tabriz, Iran

## Article history:

Received: xx xx, 2023

Accepted: xx x, 2023

ePublished: xx x, 2024

## \*Corresponding author:

Mehran Mesgari-Abbasi,  
Email: [mesgarim@tbzmed.ac.ir](mailto:mesgarim@tbzmed.ac.ir)



## Abstract

The majority of the body's organs are impacted by ambient air pollution (AAP), which is currently a serious environmental global health issue. The objective of this work was to assess the impact of ozone (O<sub>3</sub>), sulfur dioxide (SO<sub>2</sub>), and AAP on oxidative stress (OS) and lipid profile indicators in male Wistar rats. To this end, these rats were exposed for three hours each day for five weeks in control, AAP, O<sub>3</sub> (0.6 ppm), and SO<sub>2</sub> (10 ppm) groups (each containing 8 animals). Several parameters, such as total antioxidant capacity (TAC), superoxide dismutase (SOD), glutathione peroxidase (GPx) activities, low-density lipoprotein cholesterol (LDL-C), high-density lipoprotein cholesterol (HDL-C), triglyceride, and malondialdehyde (MDA), were measured on blood samples. AAP exposure on TAC ( $P=0.014$ ) and SOD ( $P=0.05$ ), MDA ( $P=0.018$ ), and HDL ( $P=0.003$ ), O<sub>3</sub> exposure on TAC ( $P<0.001$ ), and SO<sub>2</sub> exposure on blood serum TAC ( $P=0.006$ ) all had statistically significant effects. Based on the results, exposure to SO<sub>2</sub> and AAP did not significantly alter the lipid profile ( $P>0.05$ ). According to our research, exposure to O<sub>3</sub>, SO<sub>2</sub>, and AAP can increase overall antioxidant capacity by stimulating blood serum oxidative defense enzymes. Except for the enhanced effect of O<sub>3</sub> exposure on serum HDL, AAP, SO<sub>2</sub>, and O<sub>3</sub> exposures had no discernible effects on the lipid profile.

**Keywords:** Air pollution, Lipid profile, Oxidative stress, Ozone, Rats, Sulfur dioxide

**Please cite this article as follows:** Mashhadi-Abdolahi H, Darbani R, Rabet O, Golchin A, Kheirouri S, Alizadeh M, et al. Air pollution, ozone, and sulfur dioxide can affect the blood serum lipid profile and oxidative stress of male wistar rats. Avicenna J Environ Health Eng. 2024; 11(1):x-x. doi:10.34172/ajehe.5323

## 1. Introduction

Every year, exposure to air pollutants, as an environmental health problem, results in 7 million fatalities worldwide and is regarded as a risk factor for non-communicable diseases (1). Particulate matter, ozone (O<sub>3</sub>), sulfur oxides (SO<sub>2</sub>), and nitrogen oxides (NO<sub>x</sub>) are the primary constituents (2). Particulate matter smaller than 2.5 microns (PM<sub>2.5</sub>) can cause lung cancer and chronic obstructive pulmonary and heart diseases by deeply penetrating the lung and affecting the vascular and respiratory systems (3,4). The population of industrialized and developing nations is impacted by ambient air pollution (AAP). Over 91% of people on the planet are thought to reside in areas with air quality levels higher than that described in the World Health Organization's (WHO's) guidelines (5). Nevertheless, developing nations such as those in Southeast Asia and the

Western Pacific have the largest burden (6).

Evidence suggests that exposure to air pollution mostly affects the cardiovascular system, contrary to the previous assumption that it impacts respiratory diseases (7). There is evidence to support the hypothesis that oxidative stress (OS) modifies lipid metabolism and, in turn, the systemic inflammatory response (8). Triglycerides, total cholesterol (TC), low-density lipoprotein cholesterol (LDL-C), and high-density lipoprotein cholesterol (HDL-C) are factors that make up lipid profiles. Their levels are impacted by AAP, which in turn helps initiate cardiovascular events.

Conflicting findings have been found despite research on the relationship between lipid profile measures or dyslipidemia and AAP exposure (9-13). Although the lipid profile parameters were shown to be sensitive to AAP (14), some research found no correlation between AAP



and TC and HDL levels (15).

The generation of free radicals may be facilitated by exposure to environmental elements such as radiation, O<sub>3</sub>, smoke from cigarettes, and air pollution (16). An imbalance between antioxidants and free radicals is known as OS (17). Antioxidants can keep free radicals in check when they are more prevalent (18). Free radicals can harm organic molecules such as proteins, lipids, and DNA. Over time, this damage can lead to problems in human organs, such as atherosclerosis (19), heart failure (20), myocardial infarction (21), and cancer (22).

While many countries have witnessed a decrease in air pollution in recent years, industrialization, especially in those with fossil fuel resources, implies that these endeavors will not, even in the medium run, solve pollution-related issues. Thus, a more focused strategy to remove the most harmful air pollutants and maybe a way to lessen individual influences and sensitivity to air pollutants is made possible by a more comprehensive understanding of the fundamental mechanisms behind health issues brought on by air pollution (23, 24). Accordingly, this study aimed to evaluate the association between AAP, SO<sub>2</sub>, and O<sub>3</sub> on the lipid profile, including TC, HDL-C, LDL-C, and TG, as well as superoxide dismutase (SOD), catalase, glutathione peroxidase (GPx), malondialdehyde (MDA), and total antioxidant capacity (TAC) in male Wistar rats.

## 2. Materials and Methods

### 2.1. Animal Procedures

This study was conducted under approval by the Animal Research Ethics Committee located at Tabriz University of Medical Sciences under certificate No. IR.TBZMED.REC.1399.083.

Thirty-two 180-220 g weighted Wistar male rats (Pasteur Institute, Iran) were placed at 20-24 °C under 12/12-hour light/dark cycle conditions. The sample size was determined according to previous animal studies and the guidelines of the National Committee for Ethics in Biomedical Research on reducing the sample size of animal studies. The food and water were freely available. Before experimentation, the animals were acclimatized to the mentioned conditions for a week.

The animals were randomly divided into four groups (n=8 in each group) as follows:

1. The Control Group: The rats in this group only had filtered air available to them. The air filtration of this group was performed using HEPA and activated carbon filters (Panam Azma, Iran). The control group's levels of SO<sub>2</sub>, O<sub>3</sub>, and P2.5 contaminants were undetectable (25).
2. The SO<sub>2</sub> Group: For five weeks, the rats were exposed to 26 mg/m<sup>3</sup> (10 ppm) of SO<sub>2</sub> three hours a day. A 40 x 50 x 60 cm<sup>3</sup> hand-made glass enclosure with top and bottom vents. The SO<sub>2</sub> for the exposure chamber was supplied by two 50-liter SO<sub>2</sub> cylinders operating at 120 bar pressure and 1 L/Min. The gases that made up the chamber were SO<sub>2</sub> (0.002%), N<sub>2</sub> (79.5%), and

O<sub>2</sub> (20.5%). A 2-30 ppm measurement range SO<sub>2</sub> detector tube (GASTEC No. 5La) was used to check the SO<sub>2</sub> concentration in the chamber on a daily basis. The air pressure was normal, and the temperature was 22 ± 1 °C (25).

3. The Ozone group: For five weeks, three hours a day, the rats in this group were exposed to O<sub>3</sub> (1.18 ± 0.19 mg/m<sup>3</sup>, 0.6 ± 0.1 ppm) in a chamber house previously mentioned using an O<sub>3</sub> generator system (Afra Sanat, Iran). A detector (Eco Sensors, model A-21ZX-USA) was utilized to measure the O<sub>3</sub> content in the chamber daily (25).
4. The Ambient Air Pollution Group: The rats were placed in a high-traffic city square in the Tabriz city center (Abresan Square, Tabriz, East Azerbaijan, Iran) close to the air pollution recorder station for 3 hours a day for five weeks (25). The air pollutant mean concentrations for Tabriz Abresan Station during the study were extracted from Iranian AAP Monitoring System reports for Tabriz Abresan Station (Iranian Air Pollution Monitoring website, available at <http://aqms.doe.ir/Home/AQI>).

The rats were given ketamine (60 mg/kg IP) and xylazine (10 mg/kg IP) to induce unconsciousness twenty hours following the previous intervention, and blood samples were taken. The samples were put into tubes for biochemical analysis, and serum samples were isolated and kept in a freezer at -70°C (25).

### 2.2. Statistical Analyses

Data were analyzed using SPSS (Statistical Package for the Social Sciences) for Windows, version 19. The data between the groups were compared with a one-way analysis of variance, followed by Tukey's post-hoc test. The overall significance of the study was assessed at  $P < 0.05$ .

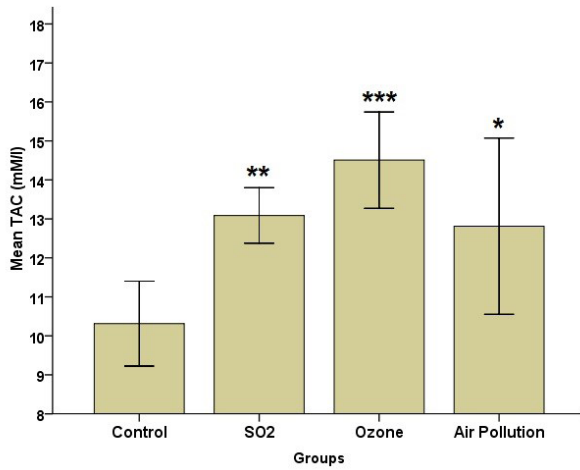
## 3. Results and Discussion

### 3.1. Air Pollution Monitoring

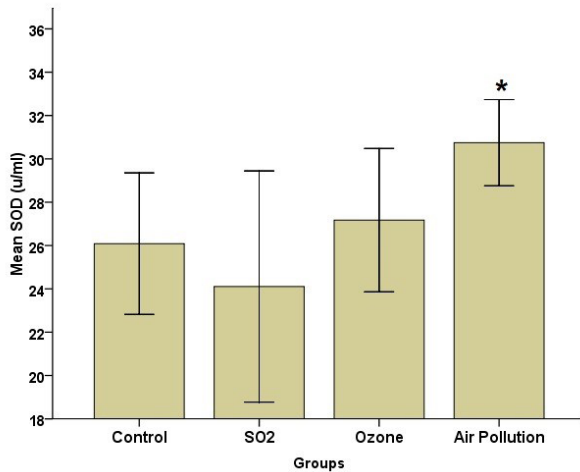
The Iranian AAP Monitoring system reported the air pollutant mean concentrations for Tabriz Abresan Station during the study. According to the results, the mean ± standard deviation (SD) of the ambient air concentration was 2.00 ± 1.17 µg/m<sup>3</sup>, 14.08 ± 10.07, and 23.96 ± 8.26 for SO<sub>2</sub>, CO, and NO<sub>2</sub>, respectively. In terms of O<sub>3</sub>, it was 49.76 ± 18.48, 28.67 ± 4.04 for PM<sub>2.5</sub>, and 21.33 ± 5.13 µg/m<sup>3</sup> for PM<sub>10</sub> (Iranian Air Pollution Monitoring website, available at <http://aqms.doe.ir/Home/AQI>).

### 3.2. Alterations in Indicators of Oxidative Stress Following Exposure to Different Gases and Ambient Air Pollution

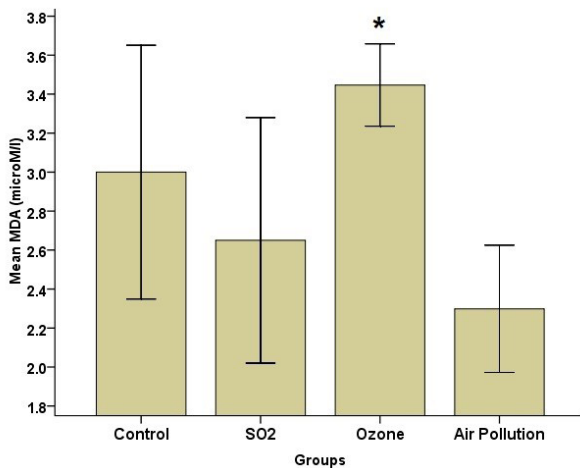
Figs. 1-3 show TAC, SOD, and MDA level changes between groups after exposure to different pollutants. As illustrated in Fig. 1, the TAC level increased in all exposure groups compared to the control. There were significant differences between the control group and SO<sub>2</sub> group



**Fig. 1.** Effects of SO<sub>2</sub>, O<sub>3</sub>, and Ambient Air Pollution Inhalation on the Blood Serum of TAC (Mean ± SD, n=8). Note. SO<sub>2</sub>: Sulfur dioxide; O<sub>3</sub>: Ozone; SD: Standard deviation; TAC: Total antioxidant capacity. \*, \*\*, and \*\*\* represent significant differences with the control group (P<0.05, P<0.01, and P<0.001 levels, respectively)



**Fig. 2.** Effects of SO<sub>2</sub>, O<sub>3</sub>, and AAP Inhalation on the Blood Serum of SOD (Mean ± SD, n=8). Note. SO<sub>2</sub>: Sulfur dioxide; O<sub>3</sub>: Ozone; SD: Standard deviation; AAP: Ambient air pollution; SOD: Superoxide dismutase. \* denotes significant differences with the SO<sub>2</sub> group (P<0.05)



**Fig. 3.** Effects of SO<sub>2</sub>, O<sub>3</sub>, and Ambient Air Pollution Inhalation on the Blood Serum of MDA (Mean ± SD, n=8). Note. SO<sub>2</sub>: Sulfur dioxide; O<sub>3</sub>: Ozone; SD: Standard deviation; AAP: Ambient air pollution; MDA: Malondialdehyde. \* indicates significant differences with the AAP group (P<0.05)

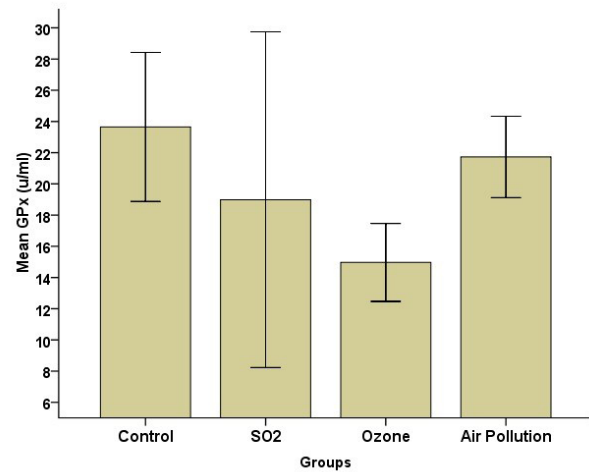
(P=0.006), O<sub>3</sub> (P<0.001), and AAP (P=0.014) groups.

According to Fig. 2, the SOD level was the highest in the AAP group, while it was the lowest in the SO<sub>2</sub> group. Group analysis, however, revealed that there was a significant difference (P=0.05) between the AAP and control groups. The MDA data (Fig. 3) indicated that the AAP group had lower MDA levels than the other groups (P=0.022). This difference, however, only matters when comparing the AAP and O<sub>3</sub> groups (P=0.018). Despite group differences in GPx, the results represented no statistically significant difference (Fig. 4).

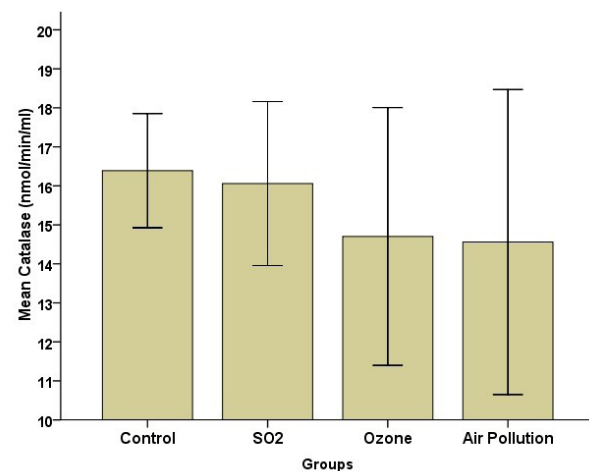
Catalase levels in the SO<sub>2</sub> and control groups were comparable, and in the other exposure groups, they were lower than in the SO<sub>2</sub> or control groups without statistical significance (Fig. 5).

### 3.3. Changes in the Blood Serum of Lipid Profiles After Ambient Air Pollution and Various Gaseous Exposure

The results related to the effects of five weeks of exposure



**Fig. 4.** Effects of SO<sub>2</sub>, O<sub>3</sub>, and Ambient Air Pollution Inhalation on the Blood Serum of GPx (Mean ± SD, n=8). Note. SO<sub>2</sub>: Sulfur dioxide; O<sub>3</sub>: Ozone; SD: Standard deviation; GPx: Glutathione peroxidase



**Fig. 5.** Effects of SO<sub>2</sub>, O<sub>3</sub>, and Ambient Air Pollution Inhalation on the Blood Serum of Catalase (Mean ± SD, n=8). Note. SO<sub>2</sub>: Sulfur dioxide; O<sub>3</sub>: Ozone; SD: Standard deviation

to SO<sub>2</sub>, O<sub>3</sub>, and AAP on the blood serum of lipid profiles are provided in Table 1. There were no discernible differences in LDL, triglyceride, or cholesterol levels across the groups. However, those in the O<sub>3</sub> group had greater HDL levels. Based on the group analysis, only the difference between the O<sub>3</sub> and control groups was significant ( $P=0.003$ ).

This study examined the association between AAP, SO<sub>2</sub>, and O<sub>3</sub> effects on the blood serum of the lipid profile and OS of male Wistar rats. The mean ambient air SO<sub>2</sub>, NO<sub>2</sub>, and O<sub>3</sub> levels during the investigation (in the AAP group) were according to the guideline values of the WHO (20 µg/m<sup>3</sup>, 40 µg/m<sup>3</sup>, and 100 µg/m<sup>3</sup>, respectively), but PM<sub>2.5</sub> and PM<sub>10</sub> concentrations exceeded the guideline values (10 µg/m<sup>3</sup> and 20 µg/m<sup>3</sup>, respectively). The results indicated that blood serum TAC and SOD significantly increased following exposure to AAP. Exposure to air pollution may have stimulated TAC and SOD levels. AAP exposure did not significantly alter the lipid profiles of the groups in our investigation. As in the report of Bernardi et al (26), the present study's AAP group had higher levels of SOD and TAC than the control group.

After fossil fuels are burned, another air pollutant called SO<sub>2</sub> is emitted into the atmosphere. Derivatives of SO<sub>2</sub> can enter the bloodstream and affect other body fluids (27). Similar to the pollutants previously described, it may involve oxidative damage to cells, tissues, and organs brought on by sulfur- and oxygen-centered free radicals produced during the oxidation of sulfite (28). Wang et al (29) did not find any linear relationship between exposure to SO<sub>2</sub> and the lipid profile of study subjects, which is compatible with our findings. In research by Wu et al (30), long-term exposure to SO<sub>2</sub> decreased the level of HDL.

Meng determined the role of SO<sub>2</sub> increment in lipid peroxidation and altered intracellular redox status in mice by administering 20 ppm (56 mg/m<sup>3</sup>) SO<sub>2</sub> to the animals for six hours per day for a week. They revealed that SO<sub>2</sub> inhalation decreased SOD and GPx compared to the control group (31). In our study, the level of catalase in SO<sub>2</sub>-exposed rats was similar to that of the control group; in addition, the levels of SOD and GPx were lower than those of the control group, which is consistent with their results. Moreover, in the present research, only the increase in TAC was statistically significant in the SO<sub>2</sub> group. This is probably because of the effects of stimuli on oxidative defense enzymes by producing OS.

The elevation of serum TAC after O<sub>3</sub> exposure was greater than that of SO<sub>2</sub>. This is possibly because of the

significant elevation in the blood serum of HDL after O<sub>3</sub> exposure. HDL has antioxidant properties and may help with TAC elevation. Neutrophilic inflammation occurs after acute exposure to O<sub>3</sub>, while oxidative pathway activation and, consequently, cell death are the outcomes of chronic exposure (32).

In this study, the blood serum level of MDA significantly increased after O<sub>3</sub> exposure. MDA is a byproduct of reactive oxygen species-induced unsaturated fatty acid oxidation, studied as a biomarker of OS. An increment in the MDA level can result from an increase in lipid peroxidation, which can be a possible result of OS induction. There is strong evidence that redox-sensitive signaling pathways mediate the pulmonary inflammatory response after exposure to a contaminant. In addition, a person's susceptibility to infection appears to be related to their antioxidant defenses (33).

The exact mechanisms of the links between air pollutants and lipid metabolism are unknown. Inhaled pollutants can induce systemic inflammation and OS, which can lead to poor lipid metabolism and oxidation. This is one proposed biological mechanism. Additionally, by decreasing the activity of DNA methyltransferases, air contaminants cause incorrect DNA methylation. Earlier studies examined the relationship between exposure to air pollutants and aberrant DNA methylation, as well as methylation in certain genes related to fat metabolism (8,34). According to a systematic review, there is a high correlation between blood lipids and air pollution in overweight and obese people (8), which is in line with the results of a previously published study (35). Air pollution is thought to be the cause of increased systemic inflammation in these situations (8).

Examining the impact of different air pollution exposures on OS indicators and lipid profiles is one of our study's strong points. To precisely determine the process underlying the adverse effects that air pollutants cause on organs, it is advised to assess these parameters on several animal model organs.

#### 4. Conclusion

According to our findings, exposure to AAP, SO<sub>2</sub>, and O<sub>3</sub> at the levels used in this investigation may increase overall antioxidant capacity via stimulating blood serum oxidative defense enzymes. Other epidemiological studies are required to evaluate the impacts of other air contaminants on biochemical markers.

**Table 1.** Changes in the Blood Serum of Lipid Profiles in Different Study Groups (Mean ±SD)

Parameters	Control (n=8)	SO <sub>2</sub> (n=8)	O <sub>3</sub> (n=8)	AAP (n=8)	P Value (Between Groups)
Triglyceride	73.20 (25.06)	84.70 (25.30)	71.67 (22.95)	70.73 (28.17)	0.710
Cholesterol	69.52 (12.21)	72.84 (19.66)	60.20 (11.17)	70.78 (11.39)	0.349
HDL	32.36 (5.30)	39.13 (5.52)	47.52 (10.35)	36.71 (7.75)	0.005
LDL	21.30 (1.96)	21.23 (0.88)	20.31 (0.97)	21.85 (1.23)	0.206

Note. SO<sub>2</sub>: Sulfur dioxide; O<sub>3</sub>: Ozone; SD: Standard deviation; HDL: High-density lipoprotein cholesterol; LDL: Low-density lipoprotein cholesterol; AAP: Ambient air pollution.

### Acknowledgments

This research was financially supported by the Drug Applied Research Center of Tabriz University of Medical Sciences, Tabriz, Iran (grant No. 62144).

### Authors' Contribution

**Conceptualization:** Sorayya Kheirouri, Mohammad Alizadeh.

**Data curation:** Roya Darbani.

**Formal analysis:** Roya Darbani.

**Funding acquisition:** Mehran Mesgari-Abbasi.

**Investigation:** Mehran Mesgari-Abbasi, Mohammad Alizadeh.

**Methodology:** Mohammad Alizadeh, Mehran Mesgari-Abbasi.

**Project administration:** Sorayya Kheirouri, Mehran Mesgari-Abbasi.

**Resources:** Sorayya Kheirouri, Mehran Mesgari-Abbasi.

**Software:** Oldouz Rabet.

**Supervision:** Mehran Mesgari-Abbasi.

**Validation:** Hossein Mashhadi-Abdolahi.

**Laboratory:** Oldouz Rabet, Roya Darbani.

**Visualization:** Asal Golchin.

**Writing—original draft:** Hossein Mashhadi-Abdolahi.

**Writing—review & editing:** Asal Golchin.

### Competing Interests

None declared.

### Data Availability Statement

The datasets generated during and/or analyzed during the current study are available from the corresponding author upon reasonable request.

### Ethical Approval

The animal procedures of the study were surveyed, approved, and monitored by the Animal Research Ethics Committee located in Tabriz University of Medical Sciences under the certificate number of IR.TBZMED.REC.1399.083) (2020-06-15).

### Funding

This research was financially supported by the Drug Applied Research Center of the Tabriz University of Medical Sciences, Tabriz, Iran (grant no. 62144).

### References

- Dhimal M, Neupane T, Lamichhane Dhimal M. Understanding linkages between environmental risk factors and noncommunicable diseases—a review. *FASEB Bioadv.* 2021;3(5):287-94. doi: [10.1096/fba.2020-00119](https://doi.org/10.1096/fba.2020-00119).
- Kampa M, Castanas E. Human health effects of air pollution. *Environ Pollut.* 2008;151(2):362-367. doi: [10.1016/j.envpol.2007.06.012](https://doi.org/10.1016/j.envpol.2007.06.012).
- Pratiwi YE, Taufik FF, Habibi J, Wibowo A. The impact of particulate matter on the respiratory system. *Jurnal Respirasi.* 2023;9(3):237-45. doi: [10.20473/jr.v9-i.3.2023.237-245](https://doi.org/10.20473/jr.v9-i.3.2023.237-245).
- Basith S, Manavalan B, Shin TH, Park CB, Lee WS, Kim J, et al. The impact of fine particulate matter 2.5 on the cardiovascular system: a review of the invisible killer. *Nanomaterials (Basel).* 2022;12(15):2656. doi: [10.3390/nano12152656](https://doi.org/10.3390/nano12152656).
- World Health Organization (WHO). WHO Global Air Quality Guidelines: Particulate Matter (PM<sub>2.5</sub> And PM<sub>10</sub>), Ozone, Nitrogen Dioxide, Sulfur Dioxide and Carbon Monoxide. WHO; 2021.
- Fatmi Z, Mahmood S, Samad Z, Wasay M. Air pollution and non-communicable diseases. *J Pak Med Assoc.* 2020;70(11):1875-7.
- Meo SA, Suraya F. Effect of environmental air pollution on cardiovascular diseases. *Eur Rev Med Pharmacol Sci.* 2015;19(24):4890-7.
- Gaio V, Roquette R, Dias CM, Nunes B. Ambient air pollution and lipid profile: systematic review and meta-analysis. *Environ Pollut.* 2019;254(Pt B):113036. doi: [10.1016/j.envpol.2019.113036](https://doi.org/10.1016/j.envpol.2019.113036).
- Yeatts K, Svendsen E, Creason J, Alexis N, Herbst M, Scott J, et al. Coarse particulate matter (PM<sub>2.5-10</sub>) affects heart rate variability, blood lipids, and circulating eosinophils in adults with asthma. *Environ Health Perspect.* 2007;115(5):709-14. doi: [10.1289/ehp.9499](https://doi.org/10.1289/ehp.9499).
- Bind MA, Peters A, Koutrakis P, Coull B, Vokonas P, Schwartz J. Quantile regression analysis of the distributional effects of air pollution on blood pressure, heart rate variability, blood lipids, and biomarkers of inflammation in elderly American men: the normative aging study. *Environ Health Perspect.* 2016;124(8):1189-98. doi: [10.1289/ehp.1510044](https://doi.org/10.1289/ehp.1510044).
- Cai Y, Hansell AL, Blangiardo M, Burton PR, de Hoogh K, Doiron D, et al. Long-term exposure to road traffic noise, ambient air pollution, and cardiovascular risk factors in the HUNT and lifelines cohorts. *Eur Heart J.* 2017;38(29):2290-6. doi: [10.1093/eurheartj/ehx263](https://doi.org/10.1093/eurheartj/ehx263).
- Yitshak Sade M, Kloog I, Liberty IF, Schwartz J, Novack V. The association between air pollution exposure and glucose and lipids levels. *J Clin Endocrinol Metab.* 2016;101(6):2460-7. doi: [10.1210/jc.2016-1378](https://doi.org/10.1210/jc.2016-1378).
- Shanley RP, Hayes RB, Cromar KR, Ito K, Gordon T, Ahn J. Particulate air pollution and clinical cardiovascular disease risk factors. *Epidemiology.* 2016;27(2):291-8. doi: [10.1097/ede.0000000000000426](https://doi.org/10.1097/ede.0000000000000426).
- Yang BY, Guo Y, Markevych I, Qian ZM, Bloom MS, Heinrich J, et al. Association of long-term exposure to ambient air pollutants with risk factors for cardiovascular disease in China. *JAMA Netw Open.* 2019;2(3):e190318. doi: [10.1001/jamanetworkopen.2019.0318](https://doi.org/10.1001/jamanetworkopen.2019.0318).
- Fioravanti S, Cesaroni G, Badaloni C, Michelozzi P, Forastiere F, Porta D. Traffic-related air pollution and childhood obesity in an Italian birth cohort. *Environ Res.* 2018;160:479-86. doi: [10.1016/j.envres.2017.10.003](https://doi.org/10.1016/j.envres.2017.10.003).
- Tripathi R, Gupta R, Sahu M, Srivastava D, Das A, Ambasta RK, et al. Free radical biology in neurological manifestations: mechanisms to therapeutics interventions. *Environ Sci Pollut Res Int.* 2022;29(41):62160-207. doi: [10.1007/s11356-021-16693-2](https://doi.org/10.1007/s11356-021-16693-2).
- Engwa GA, EnNwekegwa FN, Nkeh-Chungag BN. Free radicals, oxidative stress-related diseases and antioxidant supplementation. *Altern Ther Health Med.* 2022;28(1):114-28.
- Di Meo S, Venditti P. Evolution of the knowledge of free radicals and other oxidants. *Oxid Med Cell Longev.* 2020;2020:9829176. doi: [10.1155/2020/9829176](https://doi.org/10.1155/2020/9829176).
- Kattoor AJ, Pothineni NVK, Palagiri D, Mehta JL. Oxidative stress in atherosclerosis. *Curr Atheroscler Rep.* 2017;19(11):42. doi: [10.1007/s11883-017-0678-6](https://doi.org/10.1007/s11883-017-0678-6).
- Tsutsui H, Kinugawa S, Matsushima S. Oxidative stress and heart failure. *Am J Physiol Heart Circ Physiol.* 2011;301(6):H2181-90. doi: [10.1152/ajpheart.00554.2011](https://doi.org/10.1152/ajpheart.00554.2011).
- Ramond A, Godin-Ribuot D, Ribouot C, Totoson P, Koritchneva I, Cachot S, et al. Oxidative stress mediates cardiac infarction aggravation induced by intermittent hypoxia. *Fundam Clin Pharmacol.* 2013;27(3):252-61. doi: [10.1111/j.1472-8206.2011.01015.x](https://doi.org/10.1111/j.1472-8206.2011.01015.x).
- Klaunig JE. Oxidative stress and cancer. *Curr Pharm Des.* 2018;24(40):4771-8. doi: [10.2174/1381612825666190215121712](https://doi.org/10.2174/1381612825666190215121712).
- Pant A, Joshi RC, Sharma S, Pant K. Predictive modeling for forecasting air quality index (AQI) using time series analysis. *Avicenna J Environ Health Eng.* 2023;10(1):38-43. doi: [10.34172/ajehe.2023.5376](https://doi.org/10.34172/ajehe.2023.5376).
- Sarvi F, Nadali A, Khodadost M, Kharghani Moghaddam M, Sadeghifar M. Application of Poisson hidden Markov model to predict number of PM<sub>2.5</sub> exceedance days in Tehran during 2016-2017. *Avicenna J Environ Health Eng.* 2017;4(1):e58031. doi: [10.5812/ajehe.58031](https://doi.org/10.5812/ajehe.58031).

25. Kheirouri S, Shanebandi D, Khordadmehr M, Alizadeh M, Eskandari Vaezi F, Musapour Sultan Abad R, et al. Effects of sulfur dioxide, ozone, and ambient air pollution on lung histopathology, oxidative-stress biomarkers, and apoptosis-related gene expressions in rats. *Exp Lung Res.* 2022;48(3):137-48. doi: [10.1080/01902148.2022.2072977](https://doi.org/10.1080/01902148.2022.2072977).
26. Bernardi RB, Zanchi AC, Damaceno-Rodrigues NR, Veras MM, Nascimento Saldiva PH, Tannhauser Barros HM, et al. The impact of chronic exposure to air pollution over oxidative stress parameters and brain histology. *Environ Sci Pollut Res Int.* 2021;28(34):47407-17. doi: [10.1007/s11356-021-14023-0](https://doi.org/10.1007/s11356-021-14023-0).
27. Koenig JQ. *Health Effects of Ambient Air Pollution: How Safe is the Air We Breathe?* Springer Science & Business Media; 2000.
28. Ubuka T, Yuasa S, Ohta J, Masuoka N, Yao K, Kinuta M. Formation of sulfate from L-cysteine in rat liver mitochondria. *Acta Med Okayama.* 1990;44(2):55-64. doi: [10.18926/amo/30442](https://doi.org/10.18926/amo/30442).
29. Wang M, Zheng S, Nie Y, Weng J, Cheng N, Hu X, et al. Association between short-term exposure to air pollution and dyslipidemias among type 2 diabetic patients in northwest China: a population-based study. *Int J Environ Res Public Health.* 2018;15(4):631. doi: [10.3390/ijerph15040631](https://doi.org/10.3390/ijerph15040631).
30. Wu XM, Basu R, Malig B, Broadwin R, Ebisu K, Gold EB, et al. Association between gaseous air pollutants and inflammatory, hemostatic and lipid markers in a cohort of midlife women. *Environ Int.* 2017;107:131-9. doi: [10.1016/j.envint.2017.07.004](https://doi.org/10.1016/j.envint.2017.07.004).
31. Meng Z. Oxidative damage of sulfur dioxide on various organs of mice: sulfur dioxide is a systemic oxidative damage agent. *Inhal Toxicol.* 2003;15(2):181-95. doi: [10.1080/089583703004476](https://doi.org/10.1080/089583703004476).
32. Wiegman CH, Li F, Ryffel B, Togbe D, Chung KF. Oxidative stress in ozone-induced chronic lung inflammation and emphysema: a facet of chronic obstructive pulmonary disease. *Front Immunol.* 2020;11:1957. doi: [10.3389/fimmu.2020.01957](https://doi.org/10.3389/fimmu.2020.01957).
33. Kelly FJ. Oxidative stress: its role in air pollution and adverse health effects. *Occup Environ Med.* 2003;60(8):612-6. doi: [10.1136/oem.60.8.612](https://doi.org/10.1136/oem.60.8.612).
34. Chen Z, Salam MT, Toledo-Corral C, Watanabe RM, Xiang AH, Buchanan TA, et al. Ambient air pollutants have adverse effects on insulin and glucose homeostasis in Mexican Americans. *Diabetes Care.* 2016;39(4):547-54. doi: [10.2337/dc15-1795](https://doi.org/10.2337/dc15-1795).
35. Sørensen M, Hjortebjerg D, Eriksen KT, Ketznel M, Tjønneland A, Overvad K, et al. Exposure to long-term air pollution and road traffic noise in relation to cholesterol: a cross-sectional study. *Environ Int.* 2015;85:238-43. doi: [10.1016/j.envint.2015.09.021](https://doi.org/10.1016/j.envint.2015.09.021).